Dendritic Physical Gel: Hierarchical Self-Organization of a Peptide-Core Dendrimer to Form a Micrometer-Scale Fibrous Assembly

Woo-Dong Jang, Dong-Lin Jiang, and Takuzo Aida*

Department of Chemistry and Biotechnology Graduate School of Engineering The University of Tokyo, 7-3-1 Hongo Bunkyo-ku, Tokyo 113-8656, Japan

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Dendrimers are nanoscopic hyperbranched macromolecules with well-predictable three-dimensional shapes and potential building blocks for the construction of organized functional materials.^{1,2} Recently, self-assembly of dendrons and dendrimers has attracted great attention, where hydrogen-bonding,³ metal-ligating,⁴ van der Waals,⁵ and electrostatic interactions⁶ are utilized as the driving forces. Herein, we report the first examples of a dendritic physical gel formed by self-assembly of dipeptide-core poly(benzyl ether) dendrimers in organic solvents,^{7,8} and we highlight a "hierarchical" self-organization to form a micrometerscale fibrous assembly of nanoscale dendritic fibrils.

A series of poly(benzyl ether) dendrimers carrying *tert*butoxycarbonyl-(L)-tyrosinyl-(L)-alanine (**2**-**4**),⁹ *tert*-Boc-Tyr-(L*n*)-Ala (L*n* = *n*-layered dendritic wedge, n = 2-4, Figure 1), was synthesized by an alkaline-mediated coupling reaction of Fréchet's dendron bromides (L*n*Br; n = 2-4)¹⁰ with *tert*-Boc-(L)-Tyr-(L)-Ala¹¹ in DMF at 70 °C, and unambiguously characterized by ¹H NMR and MALDI-TOF-MS analyses.⁹ When *tert*-Boc-Tyr(L3)-Ala (**3**) was dissolved in acetonitrile (1.0 mM), and

(3) Zimmerman, S. C.; Zeng, F.-W.; Reichert, D. E. C.; Kolotuchin, S. V. Science **1996**, 271, 1095. (b) Suárez, M.; Lehn, J. M.; Zimmerman, S. C.; Skoulios, A.; Heinrich, B. J. Am. Chem. Soc. **1998**, 120, 9526. (c) Percec, V.; Ahn, C.-H.; Bera, T. K.; Ungar, G.; Yeardley, J. P. Chem. Eur. J. **1999**, 5, 1070.

(4) Newkome, G. M.; Guther, R.; Moorefield, C. N.; Cardullo, F.; Echegoyen, L.; Perezcordero, E.; Luftmann, H. Angew. Chem., Int. Ed. Engl. **1995**, 34, 2023. (b) Tomoyose, Y.; Jiang. D.-L.; Jin, R.-H.; Aida, T.; Yamashita, T.; Horie, K.; Yashima, E.; Okamoto, Y. Macromolecules **1996**, 29, 5236. (c) Issberner, J.; Vögtle, F.; Cola, L. D.; Balzani, V. Chem. Eur. J. **1997**, 3, 706. (d) Kawa, M.; Fréchet, J. M. J. Chem. Mater. **1998**, 10, 286. (e) Enomoto, M.; Aida, T. J. Am. Chem. Soc. **1999**, 121, 874.

29, 5236. (c) Issberner, J.; Vögtle, F.; Cola, L. D.; Balzani, V. Chem. Eur. J.
1997, 3, 706. (d) Kawa, M.; Fréchet, J. M. J. Chem. Mater. 1998, 10, 286. (e)
Enomoto, M.; Aida, T. J. Am. Chem. Soc. 1999, 121, 874.
(5) Kim, Y.-H. J. Am. Chem. Soc. 1992, 114, 4947. (b) Lorenz, K.; Holter,
D.; Stuhn, B.; Mülhaupt, R.; Frey, H. A. Adv. Mater. 1996, 8, 414. (c) Hudson,
S. D.; Jung, H.-T.; Percec, V.; Cho, W.-D.; Johansson, G.; Ungar, G.;
Balagurusamy, V. S. K. Science 1997, 278, 449. (d) Schenning, A. P. H. J.;
Elissen-Román, C.; Weener, J.-W.; Baars, M. W. P. L.; van der Gaast, S. J.;
Meijer, E. W. J. Am. Chem. Soc. 1998, 120, 8199. (e) Yamaguchi, N.;
Hamilton, L. M.; Gibson, H. W. Angew. Chem., Int. Ed. 1998, 37, 3275. (f)
Percec, V.; Ahn, C.-H.; Ungar, G.; Yeardley, D, J, P.; Möller, M.; Sheiko, S. Nature 1998, 391, 161.

(6) Tomioka, N.; Takasu, D.; Takahashi, T.; Aida, T. Angew. Chem., Int. Ed. 1998, 37, 1531.

(7) Terech, P.; Weiss, R. G. Chem. Rev. 1997, 97, 3133 and references therein. (b) Newkome, G. R.; Moorefield, C. N.; Baker, G. R.; Behera, R. K.; Escamillia, G. H.; Saunders, M. J. Angew. Chem., Int. Ed. Engl. 1992, 31, 917.

(8) Recent examples of gel formation behaviors of amino acid derivatives: (a) Hanabusa, K.; Okui, K.; Karaki, K.; Kimura, M.; Shirai, H. J. Colloid Interface Sci. 1997, 159, 86. (b) Mizoshita, N.; Kutsuna, T.; Hanabusa, L.; Kato, T. Chem. Commun. 1999, 781. (c) Mizoshita, N.; Hanabusa, K.; Kato, T. Adv. Mater. 1999, 11, 392.

(9) See Supporting Information.

(10) Hawker, C. J.; Fréchet, J. M. J. J. Am. Chem. Soc. **1990**, 112, 7638.

(b) L'abbé, G.; Forier, B.; Dehaen, W. Chem. Commun. 1996, 2143. (11) Sheehan, J. C.; Preston, J.; Cruikshank, P. A. J. Am. Chem. Soc. 1965, 87, 2492. (b) Chino, N.; Masui, Y.; Sakakibara, S. Peptide Chemistry, 1977; Shiba, T., Ed.; Protein Research Foundation: Japan, 1978; Vol. 12.



Figure 1. Schematic structures of dipeptide-core poly(benzyl ether) dendrimers, *tert*-Boc-Tyr(Bn)-Ala (1), *tert*-Boc-Tyr(Ln)-Ala (2–4, n = 2-4), and *tert*-Boc-Tyr(L3)-Ala-OMe (3').

the solution after sonication for 1 min was allowed to stand overnight at 20 °C, it formed a transparent gel, where about 20 000 solvent molecules are estimated to be gelled by one dendrimer molecule. The gelation was highly dependent on the concentration of 3. As the concentration of 3 was higher than 1.0 mM, the time required for the gelation was considerably shorter (e.g., 5 min at 1.9 mM), whereas no gelation was observed even after one week when the concentration of 3 was lower than 1.0 mM (e.g., 0.7 mM). The gelation also took place in other organic solvents such as acetone, ethyl acetate, CHCl₃, CH₂Cl₂, benzene, and CHCl₃/ benzene (1/1), where acetone and CHCl₃/benzene, similar to acetonitrile, were particularly effective for the gelation (critical concentrations for gelation; 4.6 and 1.9 mM, respectively). Interestingly, the gelation was also dependent on the size of the dendritic wedge: A non-dendritic benzyl-substituted dipeptide tert-Boc-Tyr(Bn)-Ala (1) and the smallest dipeptide-core dendrimer tert-Boc-Tyr(L2)-Ala (2) were highly soluble in acetonitrile and did not form gels even at a much higher concentration such as 130 mM. On the other hand, the largest dendrimer tert-Boc-Tyr(L4)-Ala (4) formed a gel in acetonitrile with a critical concentration for gelation of 1.4 mM, and also in acetone (2.2 mM) and CHCl₃/benzene (1.3 mM). Therefore, the dendritic wedge as well as the dipeptide core must be involved in the gelation process.

The gelation of dendrimers **3** and **4** is most likely induced by hydrogen-bonding interactions of the dipeptide core. In fact, upon addition of even a few drops of DMSO, the gelled samples of 3 and 4 in acetonitrile rapidly became fluid. Likewise, an inorganic salt such as LiCl, which is known to interact with amide functionalities, was found to prohibit the gelation.⁹ Infrared spectrum of a nongelled CHCl₃ solution of 4 (4.5 mM)¹² displayed relatively broad absorption bands at 3420 and 1680 cm⁻¹, characteristic of stretching vibrations of N-H and C=O bonds, respectively. On the other hand, the gelled sample of 4 in benzene/ $CHCl_3$ (1/1) (2.6 mM) showed the corresponding absorption bands at 3,320 (N-H) and 1650 cm⁻¹ (C=O), which are red-shifted by 100 and 30 cm⁻¹ from those of the nongelled CHCl₃ solution of 4. In relation to these observations, tert-Boc-Tyr(L3)-Ala-OMe (3'), an end-capped derivative of 3 having an esterified C-terminal, did not form gels but precipitated in acetonitrile and acetone, suggesting a participation of the terminal carboxylic acid functionality of the dipeptide core in the gelation process.

Cross-polarized microscopy of the gelled sample of 4 in acetonitrile showed that it contains bundles of micrometer-scale fibers. To obtain further structural information, the gelled sample of 4 in CHCl₃/benzene (1/1) was dried under reduced pressure

^{*} Author for correspondence. Telephone: +81-3-5841-7251. Fax: +81-3-5841-7310. E-mail: aida@macro.t.u-tokyo.ac.jp. (1) (a) Fréchet, J. M. J. *Science* **1994**, *263*, 1710. (b) Tomalia, D. A. *Adv.*

^{(1) (}a) Fréchet, J. M. J. Science **1994**, 263, 1710. (b) Tomalia, D. A. Adv. *Mater.* **1994**, 6, 529. (c) Fischer, M.; Vögtle, F. Angew. Chem., Int. Ed. **1999**, 38, 884.

⁽²⁾ Tomalia, D. A.; Esfand, R. Chem. Ind. 1997, 11, 416.

⁽¹²⁾ Lower than critical concentration for gelation.



Figure 2. Scanning electron microscope (SEM) images of a dried gel of 4. (A) $4000\times$, (B) $100000\times$.



Figure 3. CD spectra of *tert*-Boc-Tyr(L3)-Ala (**3**) under gelled (A: 1.2 mM, 0.01-cm quartz cell) and nongelled (B: 0.18 mM, 0.1-cm quartz cell) conditions in acetonitrile at 20 °C.

and subjected to scanning electron microscopy (SEM).¹³ From the SEM image of the dried gel (Figure 2A), the diameter of the bundled fibers was evaluated to be $1-2 \mu m$. Of further interest is the fact that each fiber consists of a bundle of much thinner fibrils with a diameter of approximately 20 nm (Figure 2B). From a computer-aided calculation of the molecular structure of **4**, it is assumed that each elementary fibril includes at least 15 dendritic wedges in the cross section. Such a hierarchical self-organization from a molecular-level association to nanoscale and further to micrometer-scale assemblies is highly interesting, since it is common to biological organisms.

The gelled sample of **3** in acetonitrile (1.2 mM) at 25 °C displayed strong circular dichrosim (CD) bands at the absorption bands of the dendritic wedge ($[\theta]_{261} = +1.06 \times 10^6$ and $[\theta]_{290} = +9.14 \times 10^5$), in addition to an enhanced CD band due to the dipeptide unit ($[\theta]_{231} = +8.26 \times 10^4$) (Figure 3A). This is in sharp contrast with the CD profile of **3** in a nongelled acetonitrile (0.18 mM,¹² Figure 3B) or dioxane (1.2 mM) solution, which displayed only a weak band due to the dipeptide unit without any detectable CD bands due to the dendritic wedge. These observations may suggest an interesting possibility that the dendritic wedges of **3** in the fibrils are arranged in a helical manner.¹⁴

Thermal behaviors of the dried gels¹⁵ were studied by differential scanning calorimetry (DSC) and temperature-variable



Figure 4. DSC profiles of dipeptide-core dendrimers; (A) *tert*-Boc-Tyr-(L2)-Ala (2), (B) *tert*-Boc-Tyr(L3)-Ala (3), and (C) *tert*-Boc-Tyr(L4)-Ala (4), in the solid state. The samples were prepared by evaporation of a nongelled acetonitrile solution of 2 and gels of 3 (15.4 mM, acetone) and 4 (10.4 mM, CHCl₃/benzene) on a piece of aluminum foil.

infrared spectroscopy. Upon heating at a rate of 10 °C min⁻¹ from 0 °C, the dried gels of 3 and 4 exhibited endothermic peaks at 112 (40.3 kJ mol⁻¹) and 122 °C (51.9 kJ mol⁻¹), respectively (Figure 4, B and C). Optical microscopy of the dried gels showed complete melting of the bundled fibers at these particular temperatures. Upon cooling at a rate of -10 °C min⁻¹, the melted samples were glassified, where the glass transition temperatures of 3 and 4 were observed at 55 and 76 °C, respectively. As described above, 2 does not form a gel in any of the above solvents. However, a solid sample of 2, prepared by evaporation of an acetonitrile solution, showed characteristic stretching vibrations of hydrogen-bonded N-H and C=O functionalities (3320 and 1650 cm⁻¹, respectively) and also displayed a melting behavior (Figure 4A) and a glass transition at much lower temperatures 74 (27.0 kJ mol-1) and 45 °C, respectively. In relation to these observations, infrared spectra of the dried samples of 2-4, upon elevating the temperature, all showed complete destruction of the hydrogen bonds at their melting temperatures. For example, the dried gel of 4 at 130 °C displayed stretching vibrational bands of the N-H and C=O bonds at 3420 and 1680 cm⁻¹, respectively, just like those observed for the nongelled CHCl₃ solution of **4**. Therefore, it is likely that the bulky dendritic wedges can stabilize the hydrogen bonds among the dipeptide cores via van der Waals interactions.

In conclusion, through the present studies on the gel-forming behaviors of 1-4 and 3' in organic solvents, we have demonstrated that large poly(benzyl ether) dendrimers carrying a dipeptide core (3, 4) are hydrogen-bonded to form fine elementary fibrils (approximately 20 nm in diameter), which are further assembled via van der Waals forces to give bundles of micrometerscale dendritic fibers. Such a hierarchical self-organization not only is interesting from a biomimetic point of view, but also may provide a new strategy for the design of micrometer-scale superstructured dendritic materials with nanometric precision.

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Supporting Information Available: Details for (1) synthetic procedures of 2-4 and 3', and (2) results of gelation experiments of **3** in the presence of LiCl (PDF). This material is available free of charge via the Internet at http://pubs.acs.org.

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⁽¹³⁾ During this drying process, the infrared spectral characteristics of the hydrogen-bonded gel remained unchanged.

⁽¹⁴⁾ Some pioneering works have been done by Percec et al.: (a) Kwon, Y.-K.; Danko, C.; Chvalun, S.; Blackwell, J.; Heck, J. A.; Percec, V. *Macromol. Symp.* **1994**, 87, 103. (b) Kwon, Y.-K.; Chvalun, S.; Schneider, A.-I.; Blackwell, J.; Percec, V.; Heck, J. A. *Macromolecules* **1994**, 27, 6129. (c) Kwon, Y.-K.; Chvalun, S. N.; Blackwell, J.; Percec, V.; Heck, J. A. *Macromolecules* **1995**, 28, 1552.

⁽¹⁵⁾ Terech, P.; Allegraud, J. J.; Garner, C. M. Langmuir 1998, 14, 3991.
(b) Terech, P. Ber. Bunsen-Ges. Phys. Chem. 1998, 102, 1630.